

# Freshwater Mollusk Biology and Conservation

## Population Demographic Data from four Populations of the Federally Endangered Rayed Bean, *Paetulunio (Villosa) fabalis* (Mollusca: Unionidae)

--Manuscript Draft--

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<b>Short Title:</b>	Population demographics of <i>Paetulunio (Villosa) fabalis</i>
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<b>Abstract:</b>	<p><i>Paetulunio fabalis</i> (formerly <i>Villosa fabalis</i>) has experienced a significant reduction in its range and is listed as endangered in both the United States and Canada. Little life history or demographic information exist for the species, but such data are critical for effective conservation. We sampled four streams in the Lake Erie and Ohio River systems of the northeastern U.S. that support populations of <i>P. fabalis</i>. We present estimates of total and relative abundance based on catch-per-unit-effort (CPUE) and quadrat sampling, the percentage of recruits, sex-specific shell length, and sex ratios for each population. We collected a total of 572 <i>P. fabalis</i> among the four streams, and the species was the fifth most abundant overall in mussel assemblages. Recruits (&lt; 20 mm shell length) were present in all streams and made up an average of 19.2% of individuals in CPUE samples and 38.2% in quadrat samples. Shell length varied among streams, but females were consistently smaller than males. Sex ratios did not differ from 1:1 at all streams. The presence of apparently large populations, vigorous recruitment, and balanced sex ratios suggest that all four streams support healthy, stable populations of <i>P. fabalis</i> that warrant protection.</p>

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2 **REGULAR ARTICLE**

3 Running head: Population demographics of *Paetulunio fabalis*

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5 **POPULATION DEMOGRAPHIC DATA FROM FOUR POPULATIONS OF THE**  
6 **FEDERALLY ENDANGERED RAYED BEAN, *PAETULUNIO (VILLOSA) FABALIS***  
7 **(MOLLUSCA: UNIONIDAE)**

8

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16

17 **ABSTRACT**

18 *Paetulunio fabalis* (formerly *Villosa fabalis*) has experienced a significant reduction in  
19 its range and is listed as endangered in both the United States and Canada. Little life  
20 history or demographic information exist for the species, but such data are critical for  
21 effective conservation. We sampled four streams in the Lake Erie and Ohio River systems  
22 of the northeastern U.S. that support populations of *P. fabalis*. For each population, we  
23 present estimates of total and relative abundance based on catch-per-unit-effort (CPUE)  
24 and quadrat sampling, the percentage of recruits, sex-specific shell length, and sex ratios.  
25 We collected a total of 572 *P. fabalis* among the four streams, and the species was the fifth-  
26 most abundant overall in mussel assemblages. Recruits (< 20 mm shell length) were present  
27 in all streams and made up an average of 19.2% of individuals in CPUE samples and  
28 38.2% in quadrat samples. Shell length varied among streams, but females were  
29 consistently smaller than males. Sex ratios did not differ from 1:1 at all streams. The  
30 presence of apparently large populations, vigorous recruitment, and balanced sex ratios  
31 suggest that all four streams support healthy, stable populations of *P. fabalis* that warrant  
32 protection.

33  
34 **KEY WORDS** – unionid, *Paetulunio fabalis*, *Villosa fabalis*, endangered, population  
35 demographics, life history

36

37

38 **INTRODUCTION**

39 Data on demographic variables, such as population size, recruitment and sex ratios, are  
40 important components for species conservation and assessing the resiliency of populations to  
41 environmental factors (Matter et al. 2013; Fonnesbeck and Dodd 2003; Connette and Semlitsch  
42 2015). Freshwater mussels (unionids) are one of the most endangered faunal groups in both  
43 North America and worldwide (Haag 2012; Graf and Cummings 2021). Demographic data are  
44 important for evaluating mussel population viability and responses of populations to stressors.  
45 For example, recruitment varies widely among species, populations and years, and can have a  
46 large effect on population growth (Haag 2012). Demographic data are lacking for most mussel  
47 populations, but they are urgently needed for conservation of rare and imperiled species.

48 Historically, the Rayed Bean, *Paetulunio fabalis* (formerly *Villosa fabalis*), was  
49 distributed throughout much of the Ohio River basin and in the Lake Erie and St. Clair drainages  
50 of the Great Lakes basin (Strayer and Jirka 1997). However, it has disappeared from much of its  
51 historical range and is now listed as endangered in both the USA and Canada (COSEWIC 2010;  
52 USFWS 2018). Little life history or population demographic information exist for the species,  
53 but such data are critical for the conservation of remaining populations.

54 We sampled four streams in the Lake Erie and Ohio River basins that support populations  
55 of *P. fabalis*. For each population, we present estimates of total and relative abundance based on  
56 catch-per-unit-effort (CPUE) and quadrat sampling, the percentage of recruits, sex-specific shell  
57 length, and sex ratios. We evaluate how these estimates differ among streams and between sexes  
58 and sampling methods. Finally, we discuss how our results inform 1) the choice of sampling  
59 methods for *P. fabalis*, and 2) an assessment of the health of these populations.

60

## 61 **METHODS**

62

### 63 **Study Area**

64 We conducted mussel surveys in four streams that support populations of *P. fabalis* (Fig.  
65 1). We surveyed one site each in Cassadaga Creek (Allegheny River drainage, Chautauqua  
66 County, New York, drainage area = 2,325 km<sup>2</sup>), Tymochtee Creek (Sandusky River drainage,  
67 Wyandot County, Ohio, 3,700 km<sup>2</sup>), and the Blanchard River (Maumee River drainage, Hancock  
68 and Hardin Counties, Ohio, 2,000 km<sup>2</sup>). We surveyed six sites in Swan Creek (Maumee River  
69 drainage, Lucas County, Ohio, 530 km<sup>2</sup>) within a 1-km section of the creek. Habitat and mussel  
70 assemblages did not differ conspicuously among these sites, and we combined data from the six  
71 sites for analysis. Sites consisted of a single stream reach (except Swan Creek) and consisted of  
72 the sample area described below.

73

### 74 **Survey Methods**

75 We conducted catch-per-unit effort (CPUE) timed searches and quadrat sampling at all  
76 sites, except the Blanchard River, where we did not conduct CPUE searches. Mussel surveys  
77 were conducted as part of environmental impact surveys associated with various construction  
78 projects and as part of a master's thesis project (Grabarkiewicz 2012). Effort and search methods  
79 varied among sites according to habitat conditions and study goals (see subsequent), but all  
80 surveys focused on detecting *P. fabalis*. We surveyed Cassadaga Creek in June 2021, Tymochtee  
81 Creek in July 2014, Blanchard River in August 2010, and Swan Creek in September 2007.

82 We conducted CPUE sampling by establishing a series of 10 × 10 m cells (100 m<sup>2</sup>) at  
83 each stream. We surveyed each cell for at least 0.83 person-hours. We surveyed 54 cells (5,400

84 m<sup>2</sup>) in Cassadaga Creek, 40 cells (4,000 m<sup>2</sup>) in Tymochtee Creek and 57 cells (5,700 m<sup>2</sup>) in  
85 Swan Creek, and total search time at each stream ranged from 33–53 person-hours (Table 1).  
86 Cells extended from bank to bank and continued upstream. We searched cells using tactile and  
87 visual methods. The latter included snorkeling, view buckets and SCUBA, depending on stream  
88 conditions. Generally, we first conducted a visual search of the cell, followed by a tactile search,  
89 during which we raked our fingers through the substrate to a depth of about 5 cm to dislodge  
90 buried mussels and we moved obstructions, such as woody debris or large rocks. After tactile  
91 searches, we conducted a final visual search to collect mussels exposed by the tactile search. We  
92 identified and measured shell length (nearest 0.1 mm) of all mussels encountered during CPUE  
93 sampling and then returned them to the stream. When possible, we also determined the sex of  
94 each *P. fabalis* based on shell morphology (COSEWIC 2010; USFWS 2018), but the sex could  
95 not be determined unambiguously for all individuals. We expressed mussel abundance estimated  
96 from CPUE sampling as number/person-hour.

97 We conducted quadrat sampling after CPUE sampling at each stream. We used a  
98 systematic sampling design with three random starts and 0.25 m<sup>2</sup> quadrats (Christman 2000;  
99 Smith et al. 2001). We excavated substrate from each quadrat by hand to a depth of  
100 approximately 15 cm, returned the substrate to the shore, and then sieved it through 6.35 mm  
101 mesh to collect all mussels in the quadrat (Vaughn et al. 1997; Obermeyer 1998; Hardison and  
102 Layzer 2001). We sampled 980 quadrats (245 m<sup>2</sup>) at Cassadaga Creek, 384 quadrats (96 m<sup>2</sup>) at  
103 Tymochtee Creek, and 450 quadrats (112.5 m<sup>2</sup>) each at Blanchard River and Swan Creek (Table  
104 1). We identified and measured shell length (nearest 0.1 mm) of all mussels encountered during  
105 quadrat sampling, determined the sex of each *P. fabalis* as described previously, and then  
106 returned all mussels to the stream. We expressed mussel abundance estimated from quadrat

107 sampling as number/m<sup>2</sup>. For both methods substrates were visually assessed while surveying at  
108 each stream.

109

## 110 **Data Analysis**

111 For all streams and both sampling methods, we calculated the percentage of the mussel  
112 assemblage represented by *P. fabalis* and all other species detected in the samples. We estimated  
113 the percentage of recruits in the population of *P. fabalis* in each stream and for both sampling  
114 methods. We identified recruits using length as a proxy for age. Our definition of a recruit was  
115 any individual < 20 mm length following Smith and Crabtree (2010).

116 We used two separate ANOVA models to examine sources of variation in length within  
117 and among populations of *P. fabalis*. We tested for differences in length between sexes and  
118 among streams using a two-factor model with interaction. For this model, we pooled length  
119 observations from CPUE and quadrat sampling. We tested for differences in length between  
120 sampling methods and among streams using a two-factor model with interaction. For this model,  
121 we pooled length observations for females and males, and we omitted the Blanchard River site  
122 because CPUE sampling was not conducted there. We tested for departures from a 1:1 sex ratio  
123 in each stream and for both sampling methods using chi-square goodness of fit tests.

124

## 125 **RESULTS**

126 We detected a total of 6,173 live individuals of 26 mussel species across all streams and  
127 both sampling methods (Table 1). We detected 15 species in both sampling methods at  
128 Cassadaga Creek, 15 and 10 species in CPUE and quadrat sampling, respectively, at Tymochtee

129 Creek, and 16 and 6 in CPUE and quadrat sampling, respectively, at Swan Creek. We detected  
130 16 species in quadrat sampling at the Blanchard River.

131 *Paetulonio fabalis* comprised a substantial percentage of the mussel assemblage in all  
132 streams but estimates of relative abundance varied among streams and sampling methods (Table  
133 1). At Cassadaga Creek, *P. fabalis* was greatly underrepresented in CPUE samples (relative  
134 abundance = 1.8%) compared with quadrat samples (22.4%). At Tymochtee Creek, estimates of  
135 *P. fabalis* relative abundance were similar for CPUE (14.1%) and quadrat samples (13.3%). At  
136 Swan Creek, *P. fabalis* was overrepresented in CPUE samples (22.0%) compared with quadrat  
137 samples (9.7%). Across all streams and sampling methods, *P. fabalis* was the fifth-most-  
138 abundant species (572 individuals) and represented 9.3% of all individuals.

139 Recruits were present in all streams, but the estimated percentage of recruits varied  
140 widely among streams and sampling methods (Table 2). The percentage of recruits was higher in  
141 quadrat samples than in CPUE samples in all streams, except Tymochtee Creek, where few *P.*  
142 *fabalis* were detected in quadrats. The percentage of recruits across streams was 3.1–49.2%  
143 (mean = 19.2%) in CPUE samples and 0.0–100.0% (mean = 38.2%) in quadrat samples. The  
144 percentage of recruits was highest for both methods in Swan Creek and lowest in Tymochtee  
145 Creek.

146 Length of *Paetulonio fabalis* varied by sex and by stream (Table 3). Sex was a significant  
147 factor in explaining variation in length, and females were smaller than males across all sites  
148 ( $F_{1,495} = 29.255$ ,  $P < 0.001$ ). Stream was also a significant factor ( $F_{3,495} = 80.165$ ,  $P < 0.001$ ), and  
149 mean length was greatest in Tymochtee Creek and lowest in Swan Creek. The sex  $\times$  stream  
150 interaction term was not significant ( $F_{3,495} = 0.943$ ,  $P = 0.4196$ ), showing that length differed  
151 between sexes in a similar way in all streams. Length did not vary by sampling method. Method

152 ( $F_{1,533} = 0.004$ ,  $P = 0.949$ ) was not a significant factor overall in explaining variation in length,  
153 but stream was ( $F_{2,533} = 17.013$ ,  $P < 0.001$ ). However, the method  $\times$  stream interaction term was  
154 significant ( $F_{2,533} = 12.657$ ,  $P < 0.001$ ), showing that the effect of method on length differed  
155 among streams. There was no evidence for a significant departure from a 1:1 sex ratio in any  
156 stream or for any sampling method (Table 3).

157

## 158 **DISCUSSION**

159 Abundance of *P. fabalis* varied among streams, but all appear to support robust and  
160 healthy populations. Density of *P. fabalis* was comparable for Cassadaga Creek, Blanchard River  
161 and Swan Creek (0.13–0.60/m<sup>2</sup>), but it was much lower at Tymochtee Creek (0.04/m<sup>2</sup>).  
162 However, total mussel density also was low at Tymochtee Creek (0.31/m<sup>2</sup>) compared with the  
163 other three streams (1.37–4.47/m<sup>2</sup>). Curiously, CPUE of *P. fabalis* at Tymotchee Creek  
164 (1.97/hour) was comparable to the other streams (1.27–4.72/hour). The discrepancy between  
165 density and CPUE estimates of *P. fabalis* at Tymochtee Creek could be a result of highly  
166 clustered aggregations of the species that were missed by quadrats but encountered by CPUE  
167 searches, which cover more area. Despite variation in abundance among streams, all of our  
168 abundance estimates are within the range reported for other surviving populations of *P. fabalis*  
169 (e.g., North Thames River = 0.016/m<sup>2</sup>; Sydenham River = 0.39-0.85/m<sup>2</sup>; Thames River =  
170 0.74/m<sup>2</sup>; French Creek = 1.5/m<sup>2</sup>; Ohio River Valley Ecosystem Team 2002; COSEWIC 2010;  
171 Smith and Crabtree 2010; Reid and Morris 2017; USFWS 2018). Notably, abundance in  
172 Cassadaga Creek, Blanchard River, and Swan Creek was similar to abundance of *P. fabalis* in  
173 the Sydenham River (0.4-0.9/m<sup>2</sup>), Ontario, which supports what is considered one of the best  
174 remaining populations of the species (COSEWIC 2010; Reid and Morris 2017; USFWS 2018).

175 Our estimates of recruitment and sex ratios further indicate that these populations are  
176 robust and healthy. We found evidence of recruitment at all sites, and recruitment was strong at  
177 Blanchard River and Swan Creek. The amount of recruitment needed to produce stable or  
178 increasing populations is unknown for *P. fabalis*, but a lack of or low recruitment is a common  
179 symptom of declining mussel populations (Haag 2012; Ćmiel et al. 2020). Population models  
180 that incorporate life span, annual survival, individual growth, and other demographic parameters  
181 are needed to better interpret recruitment in the context of population viability. Sex ratios were  
182 approximately 1:1 in all four streams, a trait shared by robust, healthy populations of *P. fabalis*  
183 in the East Sydenham and Thames rivers, Ontario, and French Creek, Pennsylvania (Metcalf-  
184 Smith et al. 1999; Smith and Crabtree 2010). Equal sex ratios often characterize large, stable and  
185 outbreeding populations, while skewed sex ratios can characterize small, isolated populations in  
186 stressful environments (Heard 1975; Haag and Staton 2003).

187 In most streams, we found *P. fabalis* in mixtures of silt, gravel, and sand substrates,  
188 similar to substrate associations reported for the species in other streams (USFWS 2018). In  
189 contrast, the substrate at Tymochtee Creek was dominated by deep silt. Silt substrate is typically  
190 considered unsuitable for *P. fabalis* (COSEWIC 2010), and this could partially explain the low  
191 abundance of *P. fabalis* and other mussel species in this stream. However, CPUE sampling  
192 revealed a substantial population of *P. fabalis*, including recruits, and species richness in  
193 Tymochtee Creek was comparable to the other streams. This finding may indicate that, at least in  
194 the Great Lakes region, silt substrate may be suitable to support stable populations of many  
195 species, including *P. fabalis*.

196 Our results corroborate the smaller size of females than males for *P. fabalis*, which is  
197 associated with other sexually dimorphic shell traits (COSEWIC 2010; USFWS 2018). Length of

198 *P. fabalis* varied slightly among streams, but mean lengths were similar to those seen in French  
199 Creek (26.9 mm) and the Sydenham and Thames rivers (27.0 and 28.0 mm, respectively)  
200 (Metcalf-Smith et al. 1999; COSEWIC 2010; Smith and Crabtree 2010).

201         Sampling methods for mussels are selected based on the goals of a study. Quadrat  
202 sampling typically provides better estimates of the abundance of recruits or small species than  
203 CPUE because small mussels can be difficult to detect by visual or tactile CPUE sampling  
204 compared with more focused quadrat sampling, particularly if substrate excavation and sieving is  
205 used (Vaughn et al. 1997; Obermeyer 1998; Smith et al. 1999). In contrast, CPUE sampling  
206 typically provides better estimates of species richness and increased detection of highly clustered  
207 mussel aggregations because more area can be searched. Our results generally support the greater  
208 efficiency of CPUE sampling for estimating species richness and greater efficiency of quadrats  
209 for detecting recruits, but they provide mixed support for other relative benefits of these  
210 methods. Because of its small size, *P. fabalis* is expected to be underrepresented in CPUE  
211 sampling compared with quadrat sampling, but we saw this at only one of three sites; at the other  
212 two sites, relative abundance was either comparable between methods or *P. fabalis* was  
213 overrepresented in CPUE samples. As discussed previously, the latter result could have been due  
214 to highly clustered aggregations of *P. fabalis* that were missed by quadrat sampling. Similarly,  
215 mean size is expected to be greater in CPUE sampling than quadrat sampling because of bias  
216 against smaller individuals by the former method. We did not observe this result consistently,  
217 and mean size across sites did not differ significantly between methods. Overall, the comparable  
218 efficiency of CPUE and quadrat sampling for detecting and characterizing length distributions of  
219 *P. fabalis* may be explained by the focus on that species in our surveys. Non-detection of *P.*  
220 *fabalis* in CPUE sampling may be more severe when study goals are focused more broadly on

221 the entire mussel assemblage. Nevertheless, our results show that use of both methods in  
222 conjunction can provide more robust assessments of abundance and size distributions (including  
223 occurrence of recruits), particularly when multiple surveys are conducted in a wide range of  
224 habitat types and conditions.

225 Our results show the existence of at least three large populations of *P. fabalis* that appear  
226 stable based on the presence of substantial recruitment. The status of the population in  
227 Tymochtee Creek is less clear, but the presence of substantial numbers of individuals, including  
228 recruits, in presumably suboptimal habitat suggests that large populations may exist in other  
229 habitats elsewhere in that stream. *Paetulonio fabalis* was reported previously from all four  
230 streams (USFWS 2018), but our site in Cassadaga Creek represents a new occurrence for the  
231 species in that stream. Although the population in Swan Creek previously was recognized as one  
232 of the largest and healthiest in the USA (USFWS 2018), little was known about the status of the  
233 populations in the other three streams. The existence of these apparently robust populations is  
234 good news for the long-term survival of *P. fabalis*, and it highlights the importance of protecting  
235 these streams. Additional demographic studies for these and other populations are needed to  
236 better assess their viability and outlook.

237

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245

## 246 LITERATURE CITED

247 Christman, M. C. 2000. A review of quadrat-based sampling of rare, geographically clustered  
248 populations. *Journal of Agricultural, Biological, and Environmental Statistics* 5:168–201.

249 Ćmiel, A. M., A. Strużyński, M. Wyrębek, A. M. Lipińska, K. Zając, and T. Zając. 2020.

250 Response of freshwater mussel recruitment to hydrological changes in a eutrophic  
251 floodplain lake. *Science of the Total Environment* 703:135467.

252 Connette, G. M., and R. D. Semlitsch. 2015. A multistate mark–recapture approach to estimating  
253 survival of PIT-tagged salamanders following timber harvest. *Journal of Applied Ecology*  
254 52:1316–1324.

255 Committee on the Status of Endangered Wildlife in Canada (COSEWIC). 2010. COSEWIC  
256 assessment and status report on the Rayed Bean *Villosa fabalis* in Canada. Ottawa,  
257 Canada: Committee on the Status of Endangered Wildlife in Canada. Available at  
258 [https://publications.gc.ca/collections/collection\\_2011/ec/CW69-14-194-2010-eng.pdf](https://publications.gc.ca/collections/collection_2011/ec/CW69-14-194-2010-eng.pdf),  
259 [accessed January 8, 2024](#).

260 Fonnesebeck, C. J., and C. K. Dodd, Jr. 2003. Estimation of flatted musk turtle (*Sternotherus*  
261 *depressus*) survival, recapture, and recovery rate during and after a disease outbreak.

262 *Journal of Herpetology* 37:602–607.

263 Grabarkiewicz, J.D. 2012. Habitat use and community structure of unionid mussels in three Lake  
264 Erie tributaries. Thesis, University of Toledo, Ohio.

265 Graf, D. L., and K. S. Cummings. 2021. A ‘big data’ approach to global freshwater mussel  
266 diversity (Bivalvia: Unionoida), with an updated checklist of genera and species. Journal  
267 of Molluscan Studies 87:1–36.

268 Haag, W. R. 2012. North American freshwater mussels: natural history, ecology, and  
269 conservation. Cambridge University Press, New York.

270 Haag, W. R., and J. L. Staton. 2003. Variation in fecundity and other reproductive traits in  
271 freshwater mussels. Freshwater Biology 48:2118–2130.

272 Hardison, B. S., and J. B. Layzer. 2001. Relations between complex hydraulics and the localized  
273 distribution of mussels in three regulated rivers. Regulated Rivers: Research and  
274 Management 17:77–88.

275 Heard, W. H. 1975. Sexuality and other aspects of reproduction in *Anodonta* (Pelecypoda:  
276 Unionidae). Malacologia 15:81–103.

277 Matter, S. F., F. Borrero, and C. Fleece. 2013. Modeling the survival and population growth of  
278 the freshwater mussel, *Lampsilis radiata luteola*. American Midland Naturalist 169:122–  
279 136.

280 Metcalfe-Smith, J. L., S. K. Staton, G. L. Mackie, and I. M. Scott. 1999. Range, population  
281 stability and environmental requirements of rare species of freshwater mussels in  
282 southern Ontario. Environment Canada, National Water Research Institute Contribution  
283 No. 99–058. Burlington, Ontario. Available at  
284 [https://www.researchgate.net/profile/Gerald-](https://www.researchgate.net/profile/Gerald-Mackie/publication/291334035_Range_Population_Stability_and_Environmental_Requirements_of_Rare_Species_of_Freshwater_Mussels_in_Southern_Ontario/links/569febf80)  
285 [Mackie/publication/291334035\\_Range\\_Population\\_Stability\\_and\\_Environmental\\_Requr-](https://www.researchgate.net/profile/Gerald-Mackie/publication/291334035_Range_Population_Stability_and_Environmental_Requirements_of_Rare_Species_of_Freshwater_Mussels_in_Southern_Ontario/links/569febf80)  
286 [ements\\_of\\_Rare\\_Species\\_of\\_Freshwater\\_Mussels\\_in\\_Southern\\_Ontario/links/569febf80](https://www.researchgate.net/profile/Gerald-Mackie/publication/291334035_Range_Population_Stability_and_Environmental_Requirements_of_Rare_Species_of_Freshwater_Mussels_in_Southern_Ontario/links/569febf80)

287 [8ae4af52546d663/Range-Population-Stability-and-Environmental-Requirements-of-](#)  
288 [Rare-Species-of-Freshwater-Mussels-in-Southern-Ontario.pdf](#), accessed January 8, 2024.

289 Obermeyer, B. K. 1998. A comparison of quadrats versus timed snorkel searches for assessing  
290 freshwater mussels. *American Midland Naturalist* 139:331–339.

291 Ohio River Valley Ecosystem Team. 2002. Status Assessment Report for the rayed bean, *Villosa*  
292 *fabalis*, occurring in the Mississippi River and Great Lakes systems Available at  
293 [https://www.researchgate.net/profile/Robert-](https://www.researchgate.net/profile/Robert-Butler/publication/275651534_Status_Assessment_Report_for_the_Rayed_Bean_Villosa_fabalis_Occurring_in_the_Mississippi_River_and_Great_Lakes_Systems_US_Fish_and_Wildlife_Service_Regions_3_4_and_5_and_Canada/links/554294d90cf23ff7168360c1/Status-Assessment-Report-for-the-Rayed-Bean-Villosa-fabalis-Occurring-in-the-Mississippi-River-and-Great-Lakes-Systems-US-Fish-and-Wildlife-Service-Regions-3-4-and-5-and-Canada.pdf)  
294 [Butler/publication/275651534\\_Status\\_Assessment\\_Report\\_for\\_the\\_Rayed\\_Bean\\_Villosa](#)  
295 [fabalis\\_Occurring\\_in\\_the\\_Mississippi\\_River\\_and\\_Great\\_Lakes\\_Systems\\_US\\_Fish\\_and](#)  
296 [Wildlife\\_Service\\_Regions\\_3\\_4\\_and\\_5\\_and\\_Canada/links/554294d90cf23ff7168360c1/](#)  
297 [Status-Assessment-Report-for-the-Rayed-Bean-Villosa-fabalis-Occurring-in-the-](#)  
298 [Mississippi-River-and-Great-Lakes-Systems-US-Fish-and-Wildlife-Service-Regions-3-4-](#)  
299 [and-5-and-Canada.pdf](#), accessed January 8, 2024.

300 Reid, S. M., and T. J. Morris. 2017. Tracking the recovery of freshwater mussel diversity in  
301 Ontario rivers: Evaluation of a quadrat-based monitoring protocol. *Diversity* 9:1–17.

302 Smith, D. R., R. F. Vilella, D. P. Lemarié, and S. von Oettingen. 1999. How much excavation is  
303 needed to monitor freshwater mussels? Pages 203–218 in R. Tankersley, D. I. Warmolts,  
304 G. T. Watters, B. J. Armitage, P. D. Johnson and R. S. Butler (editors). *Proceedings of*  
305 *the Freshwater Mollusk Symposia*. Ohio Biological Survey, Columbus, Ohio.

306 Smith, D. R., R. F. Vilella, and D. P. Lemarié. 2001. Survey protocol for assessment of  
307 endangered freshwater mussels in the Allegheny River, Pennsylvania. *Journal of the*  
308 *North American Benthological Society* 20:118–132.

309 Smith, T. A., and D. Crabtree. 2010. Freshwater mussel (Unionidae: Bivalvia) distributions and  
310 densities in French Creek, Pennsylvania. *Northeastern Naturalist* 17:387–414.

311 Strayer, D.L., and K.J. Jirka. 1997. The pearly mussels of New York State. *Memoirs of the New*  
312 *York State Museum* 26:1–113.

313 U.S. Fish and Wildlife Service (USFWS). 2018. Rayed Bean (*Villosa fabalis*) 5-year review:  
314 summary and evaluation. U.S. Fish and Wildlife Service, Midwest Region Ecological  
315 Services Field Office, Columbus, Ohio. Available at [https://ecosphere-documents-](https://ecosphere-documents-production-public.s3.amazonaws.com/sams/public_docs/species_nonpublish/2669.pdf)  
316 [production-public.s3.amazonaws.com/sams/public\\_docs/species\\_nonpublish/2669.pdf](https://ecosphere-documents-production-public.s3.amazonaws.com/sams/public_docs/species_nonpublish/2669.pdf),  
317 [accessed January 8, 2024](https://ecosphere-documents-production-public.s3.amazonaws.com/sams/public_docs/species_nonpublish/2669.pdf).

318 Vaughn, C. C., C. M. Taylor, and K. J. Eberhard. 1997. A comparison of the effectiveness of  
319 timed searches vs quadrat sampling in mussel surveys. Pages 157–162 in K. S. Cummins,  
320 A. C. Buchanan, C. A. Mayer, and T. J. Naimo (editors). *Conservation and Management*  
321 *of Freshwater Mussels II: Initiatives for the Future*. Upper Mississippi River  
322 Conservation Committee Symposium Proceedings, Rock Island, St. Louis, Missouri.

323 Table 1. Mussel abundance in four streams as estimated by catch-per-unit-effort (CPUE, number/hour) and quadrat (number/m<sup>2</sup>)  
 324 sampling. Relative abundance (percent representation in the assemblage) is given in parentheses. A dash (–) indicates a species was  
 325 not detected in sampling. CPUE sampling was not conducted at the Blanchard River.

Species	Cassadaga Creek		Tymochtee Creek		Blanchard River	Swan Creek	
	CPUE	Quadrat	CPUE	Quadrat	Quadrat	CPUE	Quadrat
<i>Actinonaias ligamentina</i>	0.02 (0.0%)	< 0.01 (0.2%)	0.03 (0.2%)	–	–	–	–
<i>Alasmidonta viridis</i>	–	–	–	–	0.02 (0.4%)	0.72 (3.3%)	0.04 (2.6%)
<i>Amblema plicata</i>	17.18 (24.0%)	0.32 (12.0%)	0.09 (0.7%)	–	0.06 (1.4%)	0.08 (0.4%)	–
<i>Anodontoides ferrussacianus</i>	–	–	0.81 (5.8%)	0.03 (10.0%)	0.07 (1.6%)	0.04 (0.2%)	–
<i>Euryntia dilatata</i>	5.04 (7.1%)	0.46 (17.0%)	–	–	1.85 (41.4%)	4.93 (22.9%)	0.85 (61.9%)
<i>Fusconaia flava</i>	–	–	2.49 (17.8%)	0.04 (13.3%)	0.33 (7.3%)	0.72 (3.3%)	0.08 (5.8%)
<i>Lampsilis cardium</i>	0.02 (0.0%)	< 0.01 (0.2%)	0.06 (0.4%)	–	0.01 (0.2%)	–	–
<i>Lampsilis ovata</i>	–	< 0.01 (0.2%)	–	–	–	–	–
<i>Lampsilis siliquoidea</i>	20.93 (29.2%)	0.38 (14.2%)	3.64 (26.0%)	0.06 (20.0%)	1.07 (24.0%)	6.45 (30.0%)	0.15 (11.0%)
<i>Lasmigona complanata</i>	–	–	–	–	0.04 (1.0%)	1.25 (5.8%)	–
<i>Lasmigona compressa</i>	0.33 (0.5%)	0.01 (0.5%)	0.30 (2.2%)	0.01 (3.3%)	–	–	–
<i>Lasmigona costata</i>	3.09 (4.3%)	0.04 (1.7%)	0.09 (0.7%)	0.01 (3.3%)	0.09 (2.0%)	0.15 (0.7%)	–
<i>Paetulunio fabalis</i>	1.27 (1.8%)	0.60 (22.4%)	1.20 (14.1%)	0.04 (13.3%)	0.29 (6.5%)	4.72 (22.0%)	0.13 (9.7%)
<i>Pleurobema sintoxia</i>	0.02 (0.0%)	< 0.01 (0.2%)	0.94 (6.7%)	–	0.05 (1.2%)	–	–
<i>Potamilus alatus</i>	–	–	–	–	–	0.09 (0.4%)	–
<i>Potamilus fragilis</i>	0.04 (0.1%)	–	–	–	–	0.15 (0.7%)	–
<i>Ptychobranchnus fasciolaris</i>	1.04 (1.5%)	0.16 (6.1%)	0.49 (3.5%)	–	0.09 (2.0%)	–	–
<i>Pyganodon grandis</i>	4.02 (5.6%)	0.09 (3.5%)	0.70 (5.0%)	0.02 (6.7%)	0.44 (9.9%)	0.45 (2.1%)	–
<i>Quadrula quadrula</i>	–	–	0.91 (6.5%)	0.04 (13.3%)	–	0.02 (0.1%)	–
<i>Sagittunio nasuta</i>	18.00 (25.1%)	0.56 (20.6%)	–	–	–	–	–
<i>Strophitus undulatus</i>	0.47 (0.7%)	0.02 (0.9%)	0.55 (3.9%)	0.04 (13.3%)	0.01 (0.2%)	0.17 (0.8%)	–
<i>Toxolasma parvum</i>	–	–	–	0.01 (3.3%)	–	–	–
<i>Truncilla truncata</i>	–	–	–	–	–	0.08 (0.4%)	–
<i>Unio merus sp.</i>	–	–	–	–	0.02 (0.4%)	–	–

<i>Utterbackia imbecillis</i>	0.11 (0.2%)	0.02 (0.6%)	–	–	–	–	–
<i>Villosa iris</i>	–	–	–	–	0.03 (0.6%)	1.49 (6.9%)	0.12 (9.0%)
Total mussel abundance	71.60	2.69	14.00	0.31	4.47	21.49	1.37
Number of species detected	15	15	15	10	16	16	6
Search time (person–hours)	45	–	33	–	–	53	–
Area sampled (m <sup>2</sup> )	–	245.0	–	96.0	112.5	–	112.5

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327 Table 2. Number of recruits observed in four populations of *Paetulunio fabalis* in catch-per-unit-effort (CPUE) and quadrat sampling.

328 Recruits were defined as individuals < 20 mm shell length. CPUE sampling was not conducted at the Blanchard River.

Site	CPUE			Quadrats			Total		
	No. of recruits	Total <i>P. fabalis</i>	Percent recruits	No. of recruits	Total <i>P. fabalis</i>	Percent recruits	No. of recruits	Total <i>P. fabalis</i>	Percent recruits
Cassadaga Creek	3	57	5.3	20	148	13.5	23	205	11.2
Tymochtee Creek	2	65	3.1	0	4	0.0	2	69	2.9
Blanchard River	-	-	-	13	33	39.4	13	33	39.4
Swan Creek	123	250	49.2	15	15	100.0	138	265	52.1
Total	128	372	34.4	48	200	24.0	176	572	30.8

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330 Table 3. Lengths and sex ratios of *Paetulunio fabalis* detected using catch-per-unit-effort (CPUE) and quadrat sampling in four  
 331 streams. Length values are means  $\pm$  SE (range).  $X^2$  and  $P$ -values are results of goodness of fit tests for departures from a 1:1 sex ratio.  
 332 CPUE sampling was not conducted at the Blanchard River.

Site	Female		Male		Unknown		Sex ratio (F:M)	$X^2$	$P$
	$N$	Length (mm)	$N$	Length (mm)	$N$	Length (mm)			
Cassadaga Creek									
CPUE	23	25.8 $\pm$ 0.5 (19-30)	26	30.3 $\pm$ 0.9 (19-38)	8	29.1 $\pm$ 2.1 (22-38)	0.9:1.0	0.18	0.67
Quadrats	81	24.2 $\pm$ 0.5 (11-35)	64	26.9 $\pm$ 0.7 (13-40)	3	24.7 $\pm$ 3.2 (19-30)	1.3:1.0	1.99	0.16
Total	104	24.6 $\pm$ 0.4 (11-35)	90	27.8 $\pm$ 0.6 (13-40)	11	27.9 $\pm$ 1.8 (19-38)	1.2:1.0	1.01	0.31
Tymochtee Creek									
CPUE	28	27.4 $\pm$ 0.5 (20-31)	36	30.2 $\pm$ 0.7 (20-38)	1	33.0 $\pm$ 0.0 (33)	0.8:1.0	1.00	0.32
Quadrats	1	27.0 $\pm$ 0.0 (27)	3	28.0 $\pm$ 2.9 (22-31)	–	–	0.3:1.0	1.00	0.32
Total	29	27.4 $\pm$ 0.4 (20-31)	39	30.1 $\pm$ 0.7 (20-38)	1	33.0 $\pm$ 0.0 (33)	1.6:1.0	1.47	0.23
Blanchard River									
Quadrats	9	21.8 $\pm$ 0.9 (19-28)	9	24.3 $\pm$ 1.1 (17-19)	15	18.1 $\pm$ 1.4 (12-29)	1:1.0	0.00	1.00
Swan Creek									
CPUE	108	18.9 $\pm$ 0.3 (13-27)	102	23.1 $\pm$ 0.4 (15-32)	40	22.3 $\pm$ 0.6 (16-31)	1.1:1.0	0.17	0.68
Quadrats	4	23.0 $\pm$ 1.2 (20-25)	9	26.3 $\pm$ 1.3 (21-33)	2	24.0 $\pm$ 1.0 (23-25)	0.4:1.0	1.92	0.17
Total	112	19.1 $\pm$ 0.3 (13-27)	111	23.3 $\pm$ 0.4 (15-33)	42	22.3 $\pm$ 0.5 (16-31)	1.0:1.0	0.00	0.95

333

334 **FIGURE LEGENDS**

335 Figure 1. Location of study sites (stars) sampled for *Paetulunio fabalis*. Inset maps show the  
336 location of the study areas in A) Ohio and B) New York, USA.

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